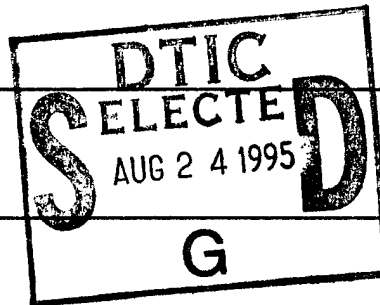


# REPORT DOCUMENTATION PAGE

Form Approved  
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE August 3, 1995		3. REPORT TYPE AND DATES COVERED Final Report (Jan 1, 1986-Dec.31, 1994)	
4. TITLE AND SUBTITLE "Biomedical Studies Using Free Electron Laser and Other Laser Systems"				5. FUNDING NUMBERS N00014-91-J-4065	
6. AUTHOR(S) J. L. Matthews, Ph.D., Principal Investigator					
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Baylor Research Institute, Dallas, TX Biomedical Engineering Dept., Univ. Texas, Austin UT-MD Anderson, Laser Biology Research Lab				8. PERFORMING ORGANIZATION REPORT NUMBER B03-832999-38	
9. SPONSORING/MONITORING AGENCY NAME(S) AND ADDRESS(ES) Department of the Navy ONR Federal Building, Room 582 300 East 8th St. Austin, TX 78701-3273				10. SPONSORING/MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES					
12a. DISTRIBUTION/AVAILABILITY STATEMENT Non-restricted distribution.				12b. DISTRIBUTION CODE	
13. ABSTRACT (Maximum 200 words) Baylor Research Institute, in collaboration with the Dept. of Biomedical Engineering at U.T. Austin and the Dept. of Laser Science at UT-M.D. Anderson hospital in Houston evaluated potential uses for high peak power pulsed and continuous wave lasers and the free-electron lasers at Duke, Stanford, and Vanderbilt for medical use. Resulting developments for potential medical use include eradication of viruses in blood banking, development of photoproducts for use in treatment of AIDS and cancer, photochemicals for non-thermal tissue welding, devices to augment thermal welding, systems for cartilage repair, combined wavelength lasers for tissue hemostasis and ablation, and a new family of photochemicals for use in tracking tumors and with potential as bonding agents in material science. Basic knowledge of laser-tissue interactions was developed.					
14. SUBJECT TERMS MFEL				15. NUMBER OF PAGES 38	
				16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT		18. SECURITY CLASSIFICATION OF THIS PAGE		19. SECURITY CLASSIFICATION OF ABSTRACT	
				20. LIMITATION OF ABSTRACT	





BAYLOR RESEARCH INSTITUTE

3812 Elm St.  
P.O. Box 710699  
Dallas, Texas 75226  
(214) 820-2687  
FAX (214) 820-4952

August 11, 1995

Department of the Navy  
Office of Naval Research  
Federal Building Room 582  
300 East 8th St.  
Austin, TX 78701-3273

Dear Sirs:

Please find enclosed requisite copies of the Final Technical Report for the MFEL program, contract number N00014-91-J-4065 at Baylor Research Institute. Thank you for your assistance.

Sincerely

J. L. Matthews, Ph.D.  
Principal Investigator

Accession For	
NTIS CRA&I	<input checked="" type="checkbox"/>
DTIC TAB	<input type="checkbox"/>
Unannounced	<input type="checkbox"/>
Justification _____	
By _____	
Distribution / _____	
Availability Codes	
Dist	Avail and / or Special
A-1	

19950818 076

DTIC QUALITY INSPECTED 5

**Final Technical Report  
Baylor Medical FEL Program**

**1. Composition of Research Group -**

The Baylor Medical Center group in Dallas teamed with the Biomedical Laser Laboratories of the University of Texas at Austin led by A. J. Welch and the Laser Biology Research Laboratory led by Steve Jacques of the University of Texas M. D. Anderson Cancer Center of Houston comprised the main core of the "Baylor" MFEL team. The major thrust of this team was the evaluation of laser-tissue interactions and the development of photochemistry as potential means of using the FEL in medical applications. We sought the three opportunities to evaluate the FEL by: 1) attempting to acquire and operate a Mark 1 Madey model of the FEL; 2) in concert with Texas A&M University and Wendel Chen of the University of Texas at Arlington and Mike Berry of Rice University we sought to build and operate an FEL at Baylor; and 3) in concert with the Boeing Corporation who developed a high peak power FEL capable of delivering high cw power in the visible spectrum. We did not win support for any of these 3 lasers so our program was focused upon using the FEL laser at Vanderbilt and at Stanford-Duke when available. In the interim periods, various commercial lasers were used to develop background information and to initiate further studies of potential uses for lasers in medicine, including high peak power pulsed lasers and conventional continuous wave lasers.

## **2. Key Projects and Accomplishments Over Duration of Program -**

### **A. Dual Use Laser Applications -**

In collaboration with Mr. D. Hulst of Lasermatic, we evaluated the potential for combined laser therapy to achieve hemostasis via coagulation concurrent with tissue ablation. This was achieved by using a broadly focused Nd/YAG laser beam with a CO<sub>2</sub> laser beam focused within the limits of the Nd/YAG beam. Since the Nd/YAG beam penetrated tissue more deeply and wider than CO<sub>2</sub> ablation beam, coagulated tissue was always in front of and lateral to the CO<sub>2</sub> cutting beam. Successful surgical ablation of highly vascular tissue such as liver and spleen was accomplished and ultimately led to FDA approved and clinical utility of the combined laser device. These data also provided the basis for future dual beam applications of the FEL.

### **B. Blood Banking Applications -**

Transmission of infectious agents is a potential risk of transfusion of whole blood or blood products. With the rapid increase in number of HIV patients came increased recognition of the importance of transfusion related transmission of infectious diseases. HIV, HBV, EBV, CMV, HTLV, hepatitis A, B, C and D are all viruses transmitted by blood as are the bacteria of Lyme's disease (*Borrelia*) and parasitic blood borne infections such as malaria, Chaga's disease (*Trypanosoma cruzi*) and *Leishmania* species as experienced in Desert Storm. We evaluated several classes of photochemicals for their potential as anti infectious compounds. Key to developing a system for blood purification was the development of chemicals that had preferential binding to virus, bacteria, or

parasite with minimal binding or uptake by important normal blood components such as RBC, platelets, and WBC. Many photochemicals activated by appropriate wavelengths of laser sources react generating highly reactive oxygen species that cause damage to adjacent cell membrane structures. We demonstrated up to 6 log<sub>10</sub> viral kill using various photochemicals such as porphyrins, merocyanines, carbocyanines, mixed ring porphyrins, sapphyrins, texaphyrins, puerpurins, psoralens, etc. and found these to be less damaging to RBC and to platelets, thus offering a potential for eradication of infectious agents in blood prior to using the blood products for transfusion. In order to optimize the kill of infectious agents and the sparing of blood cells, we synthesized two new classes of photochemicals. One of these class of new photochemicals were 1,8-naphthalimide, selected because they could act photochemically in the absence of oxygen. It was hypothesized that the damage to RBC and platelets was due to the individual exposure to reactive oxygen species generated by laser excitation. The naphthalimides were lipophylic, were concentrated in the cholesterol rich lipids of viral envelopes, and were not taken up by membranes of blood platelets, the most labile compound of the normal blood formed elements. The mechanism of viral and infectious organism inactivation was found to be cross-linking of intramembrane and transmembrane proteins. These membrane proteins change conformation upon docking with a cell to achieve infection and require conformation change for other functions. Cross-linking blocked protein change and resulted in blocking of CD<sub>4</sub> binding in case of HIV and SIV, blocked viral invasions and replication and blocked syncytial cell formation, i.e., all enveloped viruses tested (HIV, SIV, EBV, HBV, hepatitis ABCD, HTLV) were inactivated to concentration of 10 log<sub>10</sub> virus without compromising the aggregation properties, ATP, or degranulation of platelets. Platelets were functional for standard storage times.

RBC were most readily treated with porphyrin compounds. Although the second class of synthesized chemicals (texaphyrins) offered no advantages over other photochemicals for blood banking use, they have subsequently been reacted with gadolinium and are presently being employed in clinical trial as tumor imaging agents. The developed photochemicals proved effective against all enveloped viruses, against *T. cruzi* and *Leishmania* organisms, malaria of all forms and *Borrelia*. Various cw and pulsed laser sources, diode arrays, and light sources were evaluated to optimize dye/light exposure protocols. Additional support for this work was from Quadralogic Technologies, Vancouver, BC and a grant from NHLRB of NIH.

#### **C. Bone Marrow Purging -**

Some photochemicals are preferentially incorporated and/or retained by tumor cells. In treating patients who have relapsed following initial therapy, very toxic levels of chemotherapeutics are employed requiring removal of bone marrow prior to vigorous chemotherapy with return of marrow following chemotherapy. We explored the utility of treating isolated marrow cells photochemically in an effort to remove any tumor cells from marrow prior to its return to patient. Key to success of this approach is the extent to which normal marrow stem cells are spared while tumor cells are killed. Prior to photochemical studies, chemotherapeutic agents used to purge marrow of tumor cells resulted in loss of up to 80% of viable marrow stem cells. After screening numerous photochemicals, we demonstrated retention of 80% viable stem cells with 100% tumor cell kill using merocyanine compounds. This approach is now used in some marrow units. A xenon lamp with IR filtering to avoid thermal damage were found to be adequate for the task.

#### D. Use of Pre-activation of Photochemicals to Achieve Tumor Kill -

Photodynamic therapy of cancer is a relatively new approach having received FDA approval for limited use this year. The approach capitalizes on the retention of dye by tumor cells following IV administration and an incubation period of a few hours post infusion. The tumor is then irradiated with an appropriate laser source of selected wavelength in visible spectrum which results in singlet oxygen generation and subsequent lipid peroxidation and mitochondrial damage of the tumor cells. Although very successful when the tumor is accessible to the laser source, this approach is limited by tumor access in deep tissues and requires precise knowledge of the location of the tumor, i.e., metastasized cells will not be treated. We explored the possibility that photoproducts might have antitumor properties. We were led to this study by observations that the antitumor efficacy of a large battery of photochemicals of various classes did not always correlate with the capacity of the photochemical to generate singlet oxygen. Merocyanine 540 for example, was highly effective as an antitumor agent in *in vitro* studies of several cell cultures of different tumor types, but was not found to be an efficient generator of singlet oxygen. Studies on physical properties of photochemicals were done at the University of Texas in Austin in the fast kinetics spectroscopy labs directed by Dr. Tony Harriman, a collaborator on these studies. Photoproducts isolated from individual photochemicals following activation by laser light were stored at  $-80^{\circ}\text{C}$  and tested hours, days and weeks later for antitumor activity (singlet oxygen is a short-lived transient species (of the order of  $10\ \mu\text{s}$  in  $\text{H}_2\text{O}$ ) with a short diffusion path before extinction). We established varying antitumor activity with photoproducts isolated from different classes of photochemicals and significant

antitumor effects with photoproducts of merocyanine 540. Clinical use of isolated photoproducts would offer significant advantages over standard (singlet oxygen dependent) photodynamic therapy if the photoproducts were not highly toxic to normal tissues, i.e., tumors would not have to be isolated and directly exposed to laser irradiation of photoproducts achieved by prior *in vitro* radiation could be used. We named this approach "pre-activation PDT" and did an *in vitro* and *in vivo* toxicity study to ascertain whether sufficient efficacy/toxicity ratio could be realized. We also initiated attempts to isolate and ultimately characterize and synthesize the major active photoproducts so that the resultant "drug" would consist of compounds of known composition, tissue distribution, and toxicity, all necessary if photoproducts were to be quality controlled as per FDA good manufacturing guidelines.

The major photoproducts of MC540 were isolated and synthesized in collaboration with Professor Dr. B. Franck of Muenster Germany and with Dr. Gardner of Baylor University. Toxicity of these isolates was established in normal primary human cell cultures, in mice, rats, rabbits, and rhesus monkeys and found to be less than comparable chemotherapeutic agents that act principally on dividing cell populations.

A grant was obtained from Army Breast Cancer program to support the further *in vivo* evaluation of the efficacy of these photoproducts on solid tumors of breast cancer and prostate cancer tumors transplanted to nude mice. Successful eradication of transplanted tumors has been achieved with no observable short or long term toxic effects.



**E. Effects of Photoproducts on Free Virus, Viral Infected Cells, and Provirus Forms -**

Since direct PDT photochemical killing of free virus and viral infected cells was established in the blood sterilization studies, we also initiated study of the use of photoproducts as antiviral drugs. Although blood of viral infected patients could be purged of virus using an extracorporeal irradiation of patients' blood, this approach to therapy was not deemed appropriate as HIV infected patients for example, have a preponderance of infected cells and free virus resident in lymphatic and other tissues and blood circulating forms represent only a small percentage of the total viral load. Blood purging could only be a palliative means of temporarily reducing the circulating virus. However, intravenous delivery of antiviral photoproducts could at least hypothetically, circulate via blood, ECF, tissue lymph, and cerebrospinal fluid to gain access to widely dispersed infections agents. We thus investigated the *in vitro* effects of MC540 and naphthalimide photoproducts against HSV, CMV, HIV, SIV, and VSV virus. Naphthalimide photoproducts totally blocked syncytial cell formation (cell to cell infection) in SIV and HIV infected cells. However, naphthalimide photoproducts were not effective against cell-free enveloped virus at concentrations up to 100  $\mu$ m. Photoproduct inactivation of both cell-free and cellularly associated enveloped virus by treatment with photoproducts of MC540 proved to be as efficacious as Direct PDT kill of all enveloped virus. Whole animal toxicity of both MC540 and naphthalimide photoproducts was low as found in previous tumor studies. Thus a potential new antiviral compound was isolated from photoproducts of laser irradiated photochemicals. To achieve adequate quantities of "drug" for use in animal studies, a 10 x 10 ft. room was equipped with 500 fluorescent lights and mirrors to achieve

preactivation. We demonstrated efficient preactivation with a candela pulsed peak power laser and sought access to the Boeing FEL as it was the only laser source available capable of delivering visible light at adequate cw power levels with sufficient beam diameter to permit large scale volume production of photoproducts essential for phase I clinical trial activity. We sought funding for this activity in collaboration with Boeing Corporation submitting an application to the "dual use" DOD program. We were not successful in this highly competitive arena. Our need for the FEL was supplanted by our successful chemical synthesis of the photoproducts of MC540 eliminating the need for laser irradiation. Isolation and synthesis of naphthalimide photoproducts has not been achieved due to lack of funding.

**F. Use of Photochemicals and Dyes for Tissue Welding -**

Cross-linking and fusion of connective tissue fibrils, bundles and sheaths using thermal laser sources such as the medical CO<sub>2</sub> laser has been used to achieve reanastomosis of blood vessels, nerves by fusing the tunica and ventitia of vessels and the epineurium of nerve. Binding of collagen-rich tissues such as fascia, ligaments, tendons, aponeuroses, etc. is readily achieved by delivering infrared energy to the zone of fusion which results in denaturation of protein, uncoiling and fraying of uncoiled ends, loss of order and axial periodicity, and resultant melding of apposed parts. Successful bonds have been achieved, possessing adequate bond strength evidenced by tests of bond tensile strength and resistance to shear and by the fluid bearing competency of welded blood vessels.

Direct thermal welding suffers from the drawback of damage to collateral tissues unless the beam can be narrowly focused and the thermal delivery pulsed to minimize thermal diffusion from weld sites to adjacent tissue. To enhance welding resolution and reduce collateral thermal diffusion, we developed a device to deliver dyes whose absorption maximum were coincident with delivered wavelengths of visible and IR laser light sources. This development was done in collaboration with MicroFab Technologies, Inc. of Plano, Texas. The piezo electric-driven tubular jets of the device (an ink jet printer system) were used to direct dye onto tissue substrates with orientation (both spatial and temporal) of the optical axis coinciding with the pulsed dye delivery system. This system was reduced to practice and several papers have been presented employing this system for tissue welding and for ablation and shaping of both dental tissues and ear ossicles.

**G. Use of Photochemicals to Achieve Non-Thermal Tissue Bonding -**

The ideal tissue bonding system would cause no collateral damage by denaturation of protein and would achieve bonds via co-valent chemical links such that maximum tensile strength and resistance to shear would be attained where the bond is equal to that of bulk intact tissue. Using oxygen independent 1,8-naphthalimide photochemical synthesized in our labs, we have achieved covalent links between certain amino acids of collagen (lysine, hydroxylysine, tryptophan, tyrosine, methionine) and the photochemical naphthalimide dyes. By creating dyes with two reactive ends separated by carbon chains of variable length, it is possible to span interfibril distances and to covalently link each end of the dye to a collagen fibril, achieving a tissue bond that is stable against protease and collagenase attack. Electrophoretic studies of bonded proteins and

ultrastructural studies and enzyme digest studies confirm stable bonding of collagen fibrils requiring no thermal energy but rather, occurring as a consequence of photochemical reactions upon irradiation with visible light. Dye size, charge, and lipophylicity has been modified to permit bonding or welding of two tissue surfaces together ordinarily resistant to bonding because of the presence of large quantities of anionically charged proteoglycans ensheathing collagen fibrils. Three such difficult tissues bonded by this system are cartilage, joint meniscus tissue, and cornea. Support for continued development of the cartilage and meniscus bonding system was won from the American Arthritis Foundation and work at this time has been extended to clinical trials in sheep joints using naphthalimide dyes and blue semiconductor laser light. The corneal work is the subject of an SBIR application to NASA made by SY Technologies, Inc., Huntsville, AL who are producing a unique optical delivery system capable of delivering a ring of blue laser light for corneal transplant welding applications. This work was initiated in the MFEL program and currently is being done in collaboration with Dr. George Timberlake of the Ophthalmology Department of the University of Kansas Medical Center, Kansas City, MO.

## **H. Pulsed FEL IR Pumping of 1,8-naphthalimides for Crosslinking with Nucleophiles in Prostheses and Tissues -**

The photochemical cross-linking of proteins by the 3-halo-4-alkylamino-naphthalimides is a process requiring specific structural features in the protein. This is evidenced by the different light exposures at  $\lambda_{\text{abs}} = 425 \text{ nm}$  required for viral inactivation, typically  $20 \text{ J/cm}^2$ . These differences probably arise from different mechanisms involving rapid covalent reaction between the dye and electron-rich amino acid residues in viral proteins such as tryptophan as embodied in our proposed photoautomerization single electron transfer mechanism of the photoalkylation and a tentatively proposed mechanism featuring less energetically favored Michael-addition-rearomatization involving hydroxylysine and lysine residues of collagen.

One of the most important factors in our choice of the naphthalimide nucleus is its chemical stability. Substitution of the amide nitrogen by another group under normal conditions is extremely difficult. The amide linkage of 1,8-naphthalimide differs from protein amide linkages in having two carbonyl stretching frequencies; e.g., asymmetric and symmetrical stretch frequencies near  $1690$  and  $1650 \text{ cm}^{-1}$ , respectively (measured in cyclohexane for the unbrominated analog of Ed66Br  $1699 \text{ cm}^{-1}$  ( $5.886 \mu\text{M}$ ) and  $1662 \text{ cm}^{-1}$  ( $6.017 \mu\text{M}$ ), respectively). The latter band, but not the former, is near the protein amide band near  $1650 \text{ cm}^{-1}$  ( $6.06 \mu\text{M}$ ).

This difference will allow selective FEL IR excitation of the asymmetric stretch of the 1,8-naphthalimide carbonyl group in the presence of proteins, thus

potentially enhancing the reactivity of the amide group towards nucleophiles, especially of the amine-type present in collagen lysine and hydroxylysines.

We began a pilot experiment in collaboration with K.D. Straub, M.D., Ph.D. at the Duke MFEL facility in May 1993. We planned to study the potential IR-driven reaction between Ed66, the non-brominated analog of Ed66Br and aniline ( $C_2H_5NH_3$ ). At this writing, the experiment still has not been performed. The IR photoirradiation cell was designed by us and was fabricated at Duke and the visible light optics train and photomultiplier installed for real-time sampling of the kinetics of formation of the reaction product. This was to have been done by following the blue absorption shift of circa -10 nm anticipated in the 415 nm absorption peak upon substitution of the aromatic ring of aniline for the alkyl chain of the amide group. We still await IR beam availability at  $5.886\ \mu m$  wavelength following still ongoing replacement of FEL components at Duke.

Attainment of high asymmetrical stretch vibrational states should dramatically increase the polarity of the amide group. Both the increased polarity and vibrational amplitude should enhance the reaction coordinate motion for nucleophile (amine) addition to the carbonyl group in the initial reaction step leading to substitution of the amide nitrogen of the naphthalimide by the free amino nitrogen of the amino acid or protein residue. With a dimeric non-brominated 1,8-naphthalimide analog of DiEd66Br these reactions would lead to cross-linking of lysine and hydroxylysines of collagen. Thus, these reactions offer potential use of the rapid pattern flexibility MFEL in attachment of portions of collagen prostheses for transplant use during their manufacture. This flexibility in manufacture is not possible with existing technology.

**I. Laser Ablation Mechanism Studies: Laser Biology Research  
Laboratory, M.D. Anderson (Jacques *et al*)**

Our analysis has emphasized that tissue optics specify the primary zone of deposition of optical energy, called here the optical zone. The tissue absorption and scattering, the laser beam-width, and the size of pigmented structures can specify the optical zone. The laser-pulse duration can be sufficiently short that thermal diffusion is avoided and thermal energy is confined to the optical zone. The result is maximal laser-induced temperatures. The laser-pulse duration can also be sufficiently short that stress-wave propagation is avoided and stress induced by thermoelastic expansion to the optical zone is avoided. The result is maximal stress fields, which are able to propagate as stress waves to tissue sites outside the optical zone. The relationship between the optical zone and the pulse duration determines the laser-tissue interaction.

Limiting the thickness, hence volume, of the tissue optical zone in relatively non-absorbing tissues has been achieved by applying an absorbing pigment such as indocyanine green (ICG) to the surface. Our computer modeling study has disclosed four distinct phases which occur during tissue ablation upon lasing at 750-900 nm. The phases are: a) initial heating due to ICG absorption, b) evaporation with surface clamped at 100°C which desiccates surface layer, c) heating of surface after desiccation has slowed evaporation, d) rapid heating after onset of carbonization due to combination of desiccation and heating. Experiments with *ex vivo* human sclera specimens, ICG delivered with piezoelectrically driven (MicroFab, Inc.) droplets of 1% solution, and a 3-W

diode laser (2-mm dia. spot; 860 nm) demonstrated ablation and allowed comparison of theory and experiment.

The role of carbonization during laser ablation of tissue also was investigated. A dual-laser system was tested, in which a pulsed alexandrite laser thinned the carbon layer produced by a Nd:YAG laser. The thinning of the carbon layer modified the efficiency of ablation and the extent of surrounding thermal coagulation. Dual-laser irradiation offers a means of selecting between a cutting tool and a coagulation tool.

**J. Tissue Optical Property Changes In Thermal Coagulation: Laser Biology Research Laboratory, M.D. Anderson (Jacques *et al*)**

Studies have focused on relating, ultimately quantitatively, thermally-induced changes in tissue optical properties arising from laser irradiation energy input to the depths of thermal damage with a view to quantitatively monitoring damage during ablation and welding.

Thermal coagulation of albino rat skin heated *in vitro* was found to result in prominent changes of light scattering but relatively little in light absorption based on measurements using an integrating sphere spectrometer. The reduced scattering coefficients  $\mu_s(1-g)$ , gradually increase as temperatures increase from room temperature to 55°C then rapidly decrease to plateau after 70°C is reached. The differences among the  $\mu_s(1-g)$  values for the different wavelengths were greater at the lower temperatures than at higher temperatures. The absorption coefficient,  $\mu_a$ , changed very little over the test temperature range (room temperature to 90°C) and then only at higher temperatures and for longer wavelengths. The optical property changes were associated with thermally



induced light microscopic and ultrastructural changes in the dermal collagen, a major tissue component of skin.

Thermal fusion or welding of collagen-rich tissues involves the thermal denaturation of collagen which is reflected by changes in birefringence intensity in histologic sections. The weld bond between two severed edges is formed when the apposed ends of the collagen fibrils unravel during heating the re-entwine during the cooling phase. Thermal coagulation of collagen can be described as an end point of a kinetic rate process of thermal damage which is linear with time of exposure and exponential with temperature. The kinetic rate coefficients,  $A$  ( $s^{-1}$ ) and  $E$  (J/mole) in the Arrhenius formulation, have been experimentally determined for birefringence loss in rat skin collagen heated *in vitro* —  $A = 1.606 \times 10^{45}$  and  $E = 3.06 \times 10^5$ . Loss in collagen birefringence is a rare quantitative indicator of thermal damage; in this case, the structural alteration in tissue native-form collagen. The kinetic model coefficients were derived from exposure times between 600 and 6000s over the temperature range 45 to 90°C. Room temperature control specimens were also analyzed for comparison.

#### **K. Biomedical Laser Laboratories at the University of Texas at Austin, Feedback Control Mechanisms for Laser Assisted Surgery:**

Since photothermally induced alteration of collagen substructure is believed to allow tissue fusion, temperature and tissue reflectance have been investigated as markers of thermal damage.

In a previous project, a robotic system that uses transient reflectance change to control the depth and the extent of laser induced retinal lesions in rabbit models was demonstrated. In another project, controlled temperature welding was implemented to control and limit the extent of thermal damage to tissue during laser assisted photothermal welding and to achieve higher weld strengths. A prototype automated system for constant temperature vessel welding was developed and quasi-constant temperature vessel welding was performed using *in vitro* human saphenous veins. Three to five millimeter long longitudinal incisions were sealed with irradiation from a shutter controlled argon ion laser (Trimedyne, Optilase™ Contact Laser System, Santa Ana, CA). A lead selenide detector (Infrared Industries, Orlando, FL) was employed to measure tissue surface temperature and the temperature signal from the detector controlled the laser radiation by closing the laser shutter whenever a preselected control temperature was exceeded. The average bursting pressure values (ranging between 90 and 120 mmHg) of controlled temperature welds at temperatures from 100 to 120°C were 80% higher than welds at temperatures from 70 to 90°C. The prototype system was also used to perform photothermal welding of canine jejunum *in vitro*. Eight to ten millimeter long longitudinal incisions on canine jejunum were sealed with irradiation from an argon ion laser (COHERENT, Innova 100-20, Palo Alto, CA) and the surface temperature on the site of impact was monitored without feedback control. The welded incisions did not leak within the intraluminal pressure was raised to 75 mmHg. It was observed that successful seals were created at irradiances of 140 to 280 mW/mm<sup>2</sup> which produced surface temperatures above 80°C.

Currently, a surgical device for temperature feedback-controlled laser surgery is under development. This device incorporates laser delivery and a temperature

feedback sensor into one single housing. The need to coalign/cofocus the temperature sensor and the laser beam before each experiment has been eliminated. This surgical device attached to the controlled temperature system has been used to compare controlled temperature tissue welding to uncontrolled laser tissue welding on *in vitro* and *in vivo* rat intestine.

**Papers Presented at Meetings -  
Abstracts and Published Symposia**

Photosensitization of Burkitt Lymphoma (Daudi) Cells and Normal Bone Marrow Cells. S.

Pervaiz, K.S. Gulliya, J.L. Matthews, J. Fay, and R.M. Dowben. In *Proc of Southwest Section Soc Exp Biol Med*, Vol. 9, 1987.

Effect of Recombinant Interferon- $\alpha$  on Dye Mediated Laser Light Induced Photolysis of

Leukemic Cells: Implications for Bone Marrow Purging. K. Gulliya, S. Pervaiz, J.L. Matthews, J. Fay and R.M. Dowben. In *Proc of Southwest Section Soc Exp Biol Med*, Vol. 9, 1987.

Photoinactivation of Herpes Simplex Virus by Photofrin II and Light in a Flow Cell

System. F. Sogandares-Bernal, M.M. Judy, J.L. Matthews, H. Skiles and J.T. Newman. *Abstracts of Annual Meeting of the ASM*, A95, Pg. 16, 1987.

Photodynamic Inactivation of Herpes Simplex-I with Laser Irradiation. H. Skiles, F.

Sogandares-Bernal, M.M. Judy, J.L. Matthews and J.T. Newman. *Proc Biomedical Engineering VI* (Recent Development). 6th Southern Biomedical Conference, pps. 23-24, 1987.

Abstract by T.C. Chanh to American Soc Hemat 30th Annual Meeting, Dec. 1988.

Eradication of Human Small Cell Lung Carcinoma Cells by Photodynamic Therapy:

Implications for Autologous Bone Marrow Purging. K. Gulliya, J. Matthews, J. Fay, and R. Dowben. Fed of Am Soc of Exp Biol, Abstract. 1988.

Photodynamic Inactivation of Human Immunodeficiency Virus in Human Blood. J.L.

Matthews, F. Sogandares-Bernal, M. Judy, A. Marengo-Rowe, H. Skiles, J.

Leveson, T. Chanh, and J. Newman. 41st Annual Meeting, Am Assoc of Blood Banks, Abstract, October 1988.

Effect of Free Radical Quenchers on Dye-mediated Laser Light Induced Photosensitization of Leukemic Cells. K.S. Gulliya, J.L. Matthews, J.W. Fay, and R.M. Dowben. In "New Directions in Photodynamic Therapy": *Proc Int Soc Opt Eng* 847: 163-165, October, 1987, Published 1988.

Eradication of Myeloma Cells by Laser Light Induced Dye-mediated Photodynamic Therapy: Implications for Bone Marrow Purging. K. Gulliya, J.L. Matthews, R. Dowben, and J. Fay. Am. Soc. for the Adv. of Sci. Annual Meeting, Abstract, 1988.

Photodynamic Eradication of AIDS and Other Enveloped Viruses from Blood with Potential Applications for Blood Banking. A.J. Marengo-Rowe, J.L. Matthews, J.E. Leveson, J.T. Newman, F. Sogandares-Bernal, M.M. Judy, H. Skiles and T.C. Chanh. Amer. Soc. Hemat. 30th Annual Meeting, *Blood* 72:268A (Abstract #981), 1988.

Eradication of Human Small Cell Lung Carcinoma Cells by Photodynamic Therapy: Implications for Autologous Bone Marrow Purging. K. Gulliya, J. Matthews, J. Fay, and R. Dowben. Fed. of Am. Soc. of Exp. Biol., Abstract. 1988.

Photodynamic Inactivation of Human Immunodeficiency Virus in Human Blood. J.L. Matthews, F. Sogandares-Bernal, M. Judy, A. Marengo-Rowe, H. Skiles, J.

Leveson, T. Chanh and J. Newman. 41st Annual Meeting, Am Assoc of Blood Banks, Abstract, Oct. 1988.

Photodynamic Therapy of Viral Contaminants with Potential for Blood Banking Applications, Third Annual Contractors Meeting of Medical Free Electron Lasers, SDIO, Dept. of Defense, Salt Lake City, Utah, May 1988.

Photodynamic Inactivation of Human Immunodeficiency Virus in Human Blood, Am Assoc of Blood Banks 41st Annual Meeting, Oct. 1988.

Comparative Study on the Effect of Photodynamic Therapy on Normal Bone Marrow Cells and Simian Virus (SV-40) Transformed Bone Marrow Cell Line. S. Pervaiz, K. Gulliya, R. Dowben, J. Fay, J.L. Matthews and J. Singer. Fed of Am Soc of Exp Biol, Abstract, 1988.

Photodynamic Inactivation of Blood-Borne Protozoan Parasites. Seventh International Congress on Applications of Lasers and Electro-Optics. F. Sogandares-Bernal, J.L. Matthews, *et al*, San Diego, CA 1988.

Photodynamic Inactivation of Blood-Borne Enveloped Viruses. Seventh International Congress on Applications of Lasers and Electro-Optics. M. Judy, J.T. Newman, J.L. Matthews *et al*, San Diego, CA, 1988.

Photodynamic Activity Without the Necessity of Illuminating the Photosensitizer Containing Biological Targets. K.S. Gulliya, S. Pervaiz, R.M. Dowben, J.L. Matthews. *Proc SW Sec Soc Exp Biol Med.*, Abstract, 1989.

Free Radicals, Superoxide Dismutase and Neural Crest Cells. W. Davis, R. Dill, L.

Crawford, B. Nelson and J.L. Matthews. Abstract, *J Dent Res*, 1989.

The Effect of Free Radical Generating Agents on Cranial Neural Crest Cells (CNCC) in

Culture. W.L. Davis, R.E. Dill, G.R. Farmer, L. Crawford, B.C. Nelson, J.

Cooper and J.L. Matthews. Am Assoc of Anat, 120nd Annual Meeting, April 1989.

Abstract, *Anat Record*, Vol. 223, No. 4, p. 32A, 1989.

Studies of the Specificity of Xanthene Dye Binding to Mitochondria. J.R. Bunting, E.

Kamali, T.V. Phan, R.M. Dowben, and J.L. Matthews, *Proceedings of the SPIE*,

Vol. 997, 1989.

Photodynamic Inactivation of Human and Simian Immunodeficiency Viruses. T. Chanh,

G. Frenzel, G. Rappocciolo, J. Newman, M. Judy, F. Sogandares-Bernal, and J.L.

Matthews. Abstract, 73rd Annual Meeting, *Fed of Am Soc for Exp Biology J*, Vol.

3, No. 4, March 19-23, 1989.

A Novel Approach for the Inactivation of AIDS Virus by Preactivated Photoactive

Compounds. K.S. Gulliya, T.C. Chanh, J.T. Newman, S. Pervaiz, J.L. Matthews.

*J Cell Biochem* (Suppl) 14D, Abstract, 1990.

Mechanism of Antitumor and Antiviral Activity of Preactivated Photoactive Compounds.

K.S. Gulliya, S. Pervaiz, T.C. Chanh, J. Newman, and J.L. Matthews. Abstract,

*Photochem Photobiol*, 51 (suppl) 655, 1990.

*In Vivo* Toxicity of Preactivated MC540. S. Pervaiz, M. Battaglino, J.L. Matthews, and

K.S. Gulliya. *Photochem Photobiol* 51 (suppl) 65s, 1990.

Singlet Oxygen-mediated Generation of Dye-based Toxins: Implications for Systemic Therapy. K.S. Gulliya, S. Pervaiz, T.C. Chanh, J. Newman, and J.L. Matthews. Abstract. *Free Radical Biol & Med*, 9:77, 1990.

Role of Human Low Density Lipoprotein Receptors in the Increased Uptake of Merocyanine-540 by Cultured Cells. K.S. Gulliya, M. Battaglino, and J.L. Matthews. Third Biennial Meeting of the *International Photodynamic Association*, Buffalo, NY, July 170-21, P. 62, 1990.

Photodynamic Inactivation of Enveloped Viruses Using Sapphyrin, a  $22\pi$ -electron "Expanded Porphyrin": Possible Approaches to Prophylactic Blood Purification Protocols. M.M. Judy, J.T. Newman, H.L. Skiles, R. Boriack, J.L. Matthews. *SPIE, Vol 1203 Photodynamic Therapy: Mechanisms II*, p. 233-245, 1990.

Kinetic Models for Coagulation Processes: Determination of Rate Coefficients *In Vivo*. J.A. Pearce, W.F. Cheong, K. Pandit, T. McMurray, S. Thomsen. *Proc, SPIE*, Vol. 1422, p. 27-33, Lasers in Dermatology and Tissue Welding, 1991.

Thermal Coagulation of Tissues: Liver Studies Indicate a Distribution of Rate Parameters Not a Single Rate Parameter Describes the Coagulation Process. S.L. Jacques, C. Newman. X.Y. He. Proc Annual Winter Meeting of the American Society of Mechanical Engineers, Atlanta, GA, 1991.

Laser-Flash Photographic Studies of Er:YAG Ablation of Water. S.L. Jacques, G. Gofstein. *Proc SPIE*, 1525:309-312, 1991.



Optical Properties and Energy Pathways During XeCl Excimer Laser Irradiation of

Atherosclerotic Aorta. A.A. Oraevsky, S.L. Jacques, I.S. Saidi, G.H. Pettit, R.A. Sauerbrey, F.K. Tittel, P.D. Henry. 11th Ann Conf of Am Society for Laser Medicine and Surgery, San Diego, CA. *Lasers Surg Med* supplement 3, p. 5, abstract 9, 1991.

Dosimetry for Lasers and Light in Dermatology: Monte Carlo Simulations of 577-nm

Pulsed Laser Penetration into Cutaneous Vessels. S.L. Jacques, M. Keijzer. *Proc SPIE* 1422:2-13, 1991.

Inactivation of Viruses with Hematoporphyrins and Visible Light. J.L. Matthews, F.

Sogandares-Bernal, M. Judy, J.T. Newman, K.S. Gulliya and T. Chanh. *Photochem Photobiol.* Abstract, 1991.

Combined Effect of Preactivated Photofrin-II and Tamoxifen in Killing Retrofibroma

Pseudomyxoma and Breast Cancer Cells. Po-H. Chang, S. Pervaiz, M. Battaglino, J.L. Matthews, C. Clark, J. Day, J. Preskitt, D. Vanderpool and K.S. Gulliya. Abstract *Cancer Res* (in press), 1991.

Selective Toxicity of Preactivated Merocyanine Without Light. K.S. Gulliya, S. Pervaiz,

T.C. Chanh, J. Newman, and J.L. Matthews. International Photodynamic Assoc. XIV/3, Abstract, 1990.

Inactivation of Viruses with Photoactive Compounds. J.L. Matthews, F. Sogandares-

Bernal, M. Judy, K.S. Gulliya, J.T. Newman, T. Chanh, A. Marengo-Rowe. Annual Mtg of the Am Soc of Photobiol. Abstract published in *Photochem Photobiol*, P-S Song, ed, p. 42s-43s, Pergamon Press, New York, NY 1991.

Photosensitized Inactivation of Infectious Agents for Sterilization of Red Blood Cell

Concentrates and Whole Blood. M.M. Judy, J.L. Matthews, F. Sogandares-Bernal, J.T. Newman, T.C. Chanh and A. Marengo-Rowe. *Proc of the Soc of Photo-Optical Instrumentation Eng (SPIE)*, pps 129-133, Los Angeles, CA, Jan 23-24, 1992.

Soft Tissue Surgery with Combined FEL and Conventional Laser Beams. M.M. Judy,

B.L. Aronoff, J.L. Matthews, *Proc of the Soc of Photo-Optical Instrumentation Eng (SPIE)*, 1646:27-28, Los Angeles, CA, Jan 23-24, 1992.

Chemotherapy of AIDS by Photoproducts of Merocyanine 540. K.S. Gulliya, J.L.

Matthews, J.S. Allan, and T.C. Chanh. Gordon Conference, Chemotherapy of AIDS, Oxnard, CA., Abstract, March 1992.

Effects of Combined Simultaneous CO<sub>2</sub> and Nd:YAG Laser Beams on Healing of Liver.

M.M. Judy, J.L. Matthews, J.R. Goodson, B.L. Aronoff, E. Viherkoski, and D.E. Hults. Am Soc for Laser Med and Surg. Inc., Mtg., Orlando, FL, Abstract, May 1992.

Inactivation of Viruses with Photosensitizers and Visible Light. J.L. Matthews, F.

Sogandares-Bernal, M.M. Judy, J.T. Newman, T. Chanh, D.E. Lewis and R.E. Utecht. Am Chemical Soc, 24th Central Regional Meeting, Cincinnati, OH, Abstract, May 26-29, 1992.

Treatment of Platelet Concentrates. J.L. Matthews, F. Sogandares-Bernal, M.M. Judy,

J.T. Newman, T. Chanh, D.E. Lewis, and R.E. Utecht. Twentieth Annual Meeting

of the Am Soc for Photobiology, Sterilization of Blood by Light, San Marco Island, FL, Abstract, June, 1992.

Protein Cross Linking Using Lipophilic Naphthalimide Photosensitizers. J.L. Matthews, M.M. Judy, and D. Lewis. Midwest Connective Tissue Workshop, Chicago, IL, October 23-24, 1992.

Dual Laser Irradiation During Tissue Ablation: Effect of Carbon Layer Removal on Ablation and Coagulation. L. Rouben, S.L. Jacques. Proc of International Conf on Lasers '92, Soc for Opt and Quantum Electronics, Houston, TX, Dec. 1992.

Numerical Models of Photothermal Tissue Welding Processes. J.A. Pearce, T. McMurray, S. Thomsen, H. Vijverberg. Proc of International Conf on Lasers '92, Soc for Opt and Quantum Electronics, Houston, TX, Dec. 1992.

Temperature-Controlled Vessel Welding. T. Springer, A.J. Welch, and S. Thomsen. 12th Ann Conf of Am Soc for Laser Med and Surg, Lake Buena Vista, FL, *Lasers Surg Med Suppl.* 4, p. 77, Abstract 345, 1992.

Ablation of Hard Dental Tissues with the Er:YAG Laser. V.A. Vickers, S.L. Jacques, J. Schwartz, M. Motamedi, S. Rastegar, J.W. Martin. *Proc SPIE* 1646, 1992.

Laser-Flash Photography of Laser-Induced Spallation of Liquid Media. S.L. Jacques, G. Gofstein, R.S. Dingus. *Proc SPIE* 1646, 1992.

Liver Photocoagulation with Diode Laser (805 nm) vs Nd:YAG Laser (1064 nm). S.L.

Jacques, S. Rastegar, M. Motamedi, S.L. Thomsen, J. Schwartz, J. Torres, I. Mannonen. *Proc SPIE* 1646, 1992.

Kinetic Models of Tissue Fusion Processes *In Vitro*. J.A. Pearce, and S. Thomsen. *Proc SPIE*, Vol. 1643, *Low Power Laser Applications for Tissue Fusion and Biostimulation*, 1992.

Computer Simulation of Diode Ablation of Sclera Stained with Indocyanine Green. S.L.

Jacques, R.J. Erckens, J.R. Pohl, M. Motamedi. 13th Ann Mtg of the Am Soc for Lasers in Med and Surg, New Orleans, April, 1993.

Pulsed Laser Ablation of Atherosclerotic Aorta. Relative Role of Thermal and Mechanical Effects. A. Oraevsky, S. Jacques, F. Tittel, P. Henry. 13th Annual Mtg of the Am Soc for Lasers in Med and Surg., New Orleans, April, 1993.

Optical Property Changes in Thermally Coagulated Rat Skin. H. Vijverberg, R. Huang, S. Jacques, J. Schwartz, S. Thomsen. 13th Ann Mtg of the Am Soc for Lasers in Med and Surg, New Orleans, April, 1993.

The Role of Blood on Tissue Coagulation During Diode Laser Irradiation. C-T. Shen, E.D. Jansen, M. Motamedi, S.L. Jacques, S. Rastegar, A.J. Welch. 13th Annual mtg of the Am Soc for Lasers in Med and Surg, New Orleans, April, 1993.

Quantitative Morphologic Markers of Laser Thermal Injury in Cardiovascular Tissue. S. Thomsen. *Proc SPIE* Vol 1878, *Diagnostic and Therapeutic Cardiovascular Interventions*, 1993.

Changes in Optical Properties of Rat Skin During Thermal Coagulation. S. Thomsen, H. Vijverberg, R. Huang, J. Schwartz. *Proc SPIE* Vol. 1884, *Laser-Tissue Interaction IV*, 1993.

Kinetics for Birefringence Changes in Coagulated Rat Skin Collagen. J.S. Pearce, S. Thomsen, H. Vijverberg, T. McCurray. *Proc SPIE*, Vol. 1876, *Low Power Laser Applications for Laser-Tissue Fusion*, 1993.

Kinetic Models of Laser-Tissue Fusion Processes. J.A. Pearce, S. Thomsen. Proc Rocky Mountain Soc for Biomed Eng, San Antonio, TX, April, 1993.

Finite-Difference Modeling of Laser Ablation of Tissue. S.L. Jacques. *Proc SPIE*, Vol. 1884, *Laser-Tissue Interaction IV*, 1993.

How Micro is Microdissection? Laser Removal of Stratum Corneum of Skin Expose the Epidermal Battery. S.L. Jacques, F.E. Ejeckam, F. Tittel. *Proc SPIE*, Vol. 1884, *Laser-Tissue Interaction IV*, 1993.

Sidefiring Laser-Fiber Technology for Minimally Invasive Transurethral Treatment of Benign Prostate Hyperplasia. M.M. Judy, J.L. Matthews, W.W. Gardetto. Proc Soc Photo-Optical Instr Eng (SPIE) 86:1982, 1993.

Dye Microdrop Assisted Laser for Dentistry. C.J. Arcoria, C.J. Frederickson, D.J. Hayes, and M.M. Judy. *Proc of Am Soc for Lasers in Surg & Med*, 1993.

Photochemical Cross-Linking of Proteins with Visible-Light-Absorbing 1,8-

Naphthalimides. M.M. Judy, J.L. Matthews, R.L. Boriack, A. Burlacu, D.E.

Lewis, R.E. Utecht. Proc of Soc of Photo-Optical Instru Eng (SPIE), 1993.

Sidefiring Laser-fiber Technology for Minimally Invasive Transurethral Treatment of

Benign Prostatic Hyperplasia. M.M. Judy, J.L. Matthews, W.W. Gardetto, R.V.

Everett, H.A. Lahon. Proc of Soc of Photo-Opt Instru Eng (SPIE), 1993.

Cell Death and DNA Fragmentation Induced by Novel Compounds Merocil and

Merodantoin. K.S. Gulliya, B. Franck, U. Schneider, R.K. Sharma, L. Arnold, and

J.L. Matthews. Abstract, Am Assoc for Cancer Research, Inc., 1993.

Alteration of the G Protein of Vesicular Stomatitis Virus During Photoinactivation. H.L.

Skiles, Y. Yang, J.L. Matthews, J. T. Newman, D.E. Lewis, R.E. Utecht, S.T.

Nichol. J.J. Holland, M.M.Judy. Abstract, 93rd Gen Mtg, Am Soc for Microbiol,

Atlanta, GA, 1993.

MicroJet-Assisted Laser Tools for Surgery and Dentistry. D.B. Wallace, C.J. Arcoria, M.

Motamedi, M. Judy, C.J. Frederickson. Soc for Photo-Opt Instrumentation and Eng.

2128, Los Angeles, January, 1994.

Gel Electrophoretic Studies of Photochemical Cross-linking of Type I Collagen with

Brominated 1,8-naphthalimide Dyes and Visible Light. Soc for Photo-Opt

Instrument. Eng, Los Angeles, 2128:506-508, 1994.

## Publications

### Photodynamic Therapy of Viral Contaminants with Potential for Blood Banking

Applications. J.L. Matthews, J.T. Newman, F. Sogandares-Bernal, M.M. Judy, H. Skiles, J.E. Leveson, A.J. Marengo-Rowe, and T.C. Chanh. *Transfusion*, Vol. 28, No. 1, pp. 81-83, 1988.

### Photosensitization of Leukemic Cells and Normal Bone Marrow Cells by 514 nm Laser

Light and Effects of Laser Light on Migration Inhibition and Lymphokine Response. K. Gulliyya and M. Matthews. *Cell Bio International Reports* 12:305-312, 1988.

### Increased Survival of Normal cells During Laser Photodynamic Therapy: Implications for

*ex vivo* Autologous Bone Marrow Purging. K.S. Gulliyya, J.L. Matthews, J.W. Fay and R.M. Dowben. *Life Sci*, Vol. 112, pp. 2651-2656, 1988.

### Is Transformed Bone Marrow Cell Line a Substitute for Normal Bone Marrow Cells in

Laser Photoradiation Therapy for Bone Marrow Purging? S. Pervaiz, K.S. Gulliyya, R.M. Dowben and J.L. Matthews. *Baylor University Medical Center Proceedings (Dallas)*, Vol. 1, No. 4, pps. 35-45, October 1988.

### Elimination of Leukemic Cells by Laser Photodynamic Therapy. K. S. Gulliyya, J.W. Fay,

R.M. Dowben, S. Berkholder, and J.L. Matthews. *Cancer Chemo & Pharmacol*, Vol 22, pps. 211-214, 1988.

Photosensitization of Leukemic Cells and Normal Bone Marrow Cells by 514 nm Laser

Light and Effects of Laser Light on Migration Inhibition and Lymphokine Response.

K. Gulliya and J. Matthews. *Cell Bio Int'l Reports*, 12:305-312, 1988.

Inactivation of Tumors and Viruses Via Efficient Photoisomerization. J. Davilla, K.

Gulliya and A. Harriman. *J Chem Soc Chem Commun*. 30:1215-1216, 1989.

An *in vitro* Model of Autologous Bone Marrow Purging for Multiple Myeloma in Lung

Carcinoma Cells by Laser Photoradiation Therapy. *The Cancer J* 2:378-382, 1989.

Elimination of Clonogenic Tumor Cells from HL-60, Daudi, and U-937 Cell Lines by

Laser Photoradiation Therapy: Implications for Autologous Bone Marrow Purging.

K. Gulliya, S. Pervaiz. *Blood* 73:1059-1065, 1989.

Photoinactivation of Human Immunodeficiency Virus in Blood. J.L. Matthews, F.

Sogandares-Bernal, M.M. Judy, A.J. Marengo-Rowe, J.E. Leveson, H. Skiles,

T.C. Chanh, and J.T. Newman. *Transfusion*.28:81. 1988.

Inactivation of AIDS Virus by Preactivated Photoactive Compounds. K.S. Gulliya, F.

Chanh, J. Newman, S. Pervaiz, and J. L. Matthews. Submitted to *AIDS Research &*

*Human Retrovirus*. 1989.

The Immunohistochemical Localization of Superoxide Dismutase Activity in the Avian

Epiphyseal Growth Plate. W.L. Davis, M. Kipnis, K. Shibata, G.R. Farmer, E.

Cortinas, J.L. Matthews and D.B.P. Goodman. *Histochem J* 21:210-215, 1989.



Photodynamic Inactivation of Simian Immunodeficiency Virus. T.C. Chanh, J.S. Allan, J.L. Matthews, F. Sogandares-Bernal, M.M. Judy, H. Skiles, J. Leveson, A. Marengo-Rowe and J.T. Newman. *J of Virological Meth*, 26:125-131, 1989.

Chaga's Disease in Texas. F. Sogandares-Bernal, J.L. Matthews, M.V. Dennis, and M.M. Judy. *Baylor University Medical Center Proceedings*, Vol. 2, No. 4, pps. 3-14, October, 1989.

Protection of NIH 3T3 Cells from Infection by Trypomastigotes and Sphaeromastigotes of *Trypanosoma cruzi*, Telahuen Strain, by Porphyrins in the Presence and Absence of Light (630 and 690 nm). M.V. Dennis, M.M. Judy, J.L. Matthews and F. Sogandares-Bernal. *J of Parasitol*. 75(6):970-976, 1989.

Protective Qualities of Mitochondrial and Cytosolic Fluorescent Dyes Against *in vitro* and *in vivo* Infection by the Telahuen Strain of *Trypanosoma cruzi*. M.V. Dennis, M.M. Judy, J.L. Matthews, and F. Sogandares-Bernal. *J Parasitol* 76(2)171-176, 1990.

The Mechanism of LDL-Mediated Increased Uptake of Merocyanine 540 by HL-60 Cells. K. Gulliya, J. Davilla, A. Harriman. *Cancer J* 3:360-365, 1990.

Breast Cancer and Laser Photoradiation Therapy: an *in vitro* Model for Autologous Bone Marrow Purging. K. Gulliya, M. Battaglino and L. Matthews. Bone Marrow Purging & Processing. Alan R. Liss, Pub. NY, pp. 103-107, 1990.

Biomedical Applications of Ink-Jet Technology, C.J. Frederickson, D. Wallace, D.J. Hayes. *Proc for the Houston Soc for Biomed Eng*. 1991.

Tumor Cell Specific Dark Cytotoxicity of Preactivated Merocyanine 540: Implications for Systemic Therapy Without Light. K.S. Gulliya, S. Pervaiz, R.M. Dowben, and J.L. Matthews. *Photochem Photobio.* 51:831-838, 1990.

Preactivation — A Novel Anti-Tumor and Antiviral Approach. K.S. Gulliya, T. Chanh, J. Newman, S. Pervaiz, and J.L. Matthews. *Eur J Cancer*, Vol 26(4):551-553, 1990.

Mechanism of Antitumor and Antiviral Activity of Preactivated Photoactive Compounds. K.S. Gulliya, S. Pervaiz, T.C. Chanh, J. Newman and J.L. Matthews. *Photochem and Photobiol.* 51(suppl)655, 1990.

*In Vivo* Toxicity of Preactivated MC540. S. Pervaiz, M. Battaglino, J.L. Matthews, and K.S. Gulliya. *Photochem Photobiol* 51(suppl)67s, 1990.

Merocyanine 540 and Photofrin II as Photosensitizers for *in vitro* Killing of Duck Hepatitis B Virus and Human Hepatoma Cells. T.I. Lin, Y.S. Shein, M.C. Kao. *SPIE 2078, Photodynamic Therapy of Cancer. J Jori Ed.* 1990.

Singlet Oxygen-mediated Generation of Dye-based Toxin. Implications for Systemic Therapy. K.S. Gulliya, S. Pervaiz, T.C. Chanh, J. Newman, and J.L. Matthews. *Abstract Free Radical Biol & Med* 9:77, 1990.

*In vitro* Photodynamic Inactivation of Herpes Simplex Virus with Sapphyrins: 22 $\pi$ -Electron Porphyrin-like Macrocytes. M.M. Judy, J.L. Matthews, J.T. Newman, H.L. Skiles, R.L. Boriack, J.L. Sessler, M. Cyr, B.G. Maiya, and S.T. Nichol. *Photochem Photobiol* 53:1,101-107, 1991.

Synergy between Preactivated Photofrin-II and Tamoxifen in Killing Retrofibroma

Pseudomyxoma and Breast Cancer Cells. Po-H. Chang, S. Pervaiz, M. Battaglino, J.L. Matthews, C. Clark, J. Day, J. Preskitt, D. Vanderpool, and K.S. Gulliya. *Eur J Cancer*. 27(8):1034-1039, 1991.

Preliminary Studies of Photoinactivation of Human Immunodeficiency Virus in Blood.

J.L. Matthews, F. Sogandares-Bernal, M.M. Judy, A.J. Marengo-Rowe, J.E. Leveson, H. Skiles, J.T. Newman, and T.C.Chanh. *Transfusion* 31(7):636-641, 1991.

Combined Effect of Preactivated Photofrin-II and Tamoxifen in Killing Retrofibroma

Pseudomyxoma and Breast Cancer Cells. Po-H. Chang, S. Pervaiz, M. Battaglino, J.L. Matthews, C. Clark, J. Day, J.Preskitt, D. Vanderpool and K.S. Gulliya. *Cancer Research* 32:339, 1991.

Pathologic Analysis of Photothermal and Photomechanical Effects of Laser-tissue

Interactions. S.L. Thomsen. *Photochem Photobiol*. 53:825-35.1991

Mid-infrared Laser Ablation of Stratum Corneum Enhances *in vitro* Percutaneous Transport

of Drugs. J.S. Nelson, J.L. McCullough, T.C. Glenn, W.H. Wright, L.L. Liaw, S.L. Jacques. *J Invest Dermatol* 97:874-879, 1991.

Preactivated Merocyanine 540 Inactivates HIV-1 and SIV: Potential Therapeutic and Blood

Banking Applications. T.C. Chanh, J.S. Allan, S. Pervaiz, J.L. Matthews, S.R. Trevino, and K.S. Gulliya. *J of AIDS*. 5:188-195, 1992.

Light Fluence Required for Instantaneous Photodynamic Immobilization of Individual

Trypomastigotes of *Trypanosoma cruzi* by Photofrin II. F. Sogandares-Bernal, R.H. Hurst, R.L. Boriack, M.M. Judy, J.L. Matthews and D.W. Ussery. *Lasers in Life Sciences*, 4(4):219-224, 1992.

Laser-Tissue interactions: photochemical, photothermal, and photomechanical. S. Jacques. *Surgical Clinics*, 72:531-558, 1992.

XeCl Laser Ablation of Atherosclerotic Aorta: Optical Properties and Energy Pathways. A. Oraevsky, S.L. Jacques, G.H. Petit, I.S. Saidi, R.A. Sauerbrey, F.K. Tittel, P.D. Henry. *Lasers Surg Med* 12:585-597, 1992.

Rate Process Analysis of Thermal Damage. J.A. Pearce, S. Thomsen. Optical-Thermal Response of Laser Irradiated Tissue, eds. M.J.C. van Gemert and A.J. Welch, Plenum Press. 1992.

Inactivation of Viruses with Photoactive Compounds. J.L. Matthews, F. Sogandares-Bernal, M. Judy, K. Gulliya, J. Newman, T. Chanh, A. Marengo-Rowe. *Blood Cells*. 18:75-89, 1992.

Thermal Effects in Tissues from Combined Simultaneous Coaxial CO<sub>2</sub> and Nd:YAG Laser Beams. M.M. Judy, J.L. Matthews, J.R. Goodson, D.F. Hults, E. Viherkoski, B.L. Aronoff. *Lasers in Surg and Med*, 12(2):222-230, 1992.

Preactivation: A New Concept for Generation of Photoproducts for Potential Therapeutic Applications. K.S. Gulliya, T. Chanh, A. Harriman, B.L. Aronoff, and J.L. Matthews. *Sem in Surg Onc* 8:250-253, 1992.

Protein Damage by Photoproducts of Merocyanine 540. S. Pervaiz, A. Harriman, and K. Gulliya. *Free Rad Biol & Med* 12:389-396, 1992.

Photosensitizers for the Photodynamic Therapy: Results & Applications. B. Frank, U. Shreider, D. Schroder, K. Gulliya and L. Matthews. *Biologic Effects of Light*. Waller de Gryter, Berlin. 330, 1992.

Influx and Efflux Kinetics of Cationic Dye Binding to Respiring Mitochondria. J.R. Bunting. *Biophys Chem*, 42:162-175, 1992.

Neutralization of HIV-1 and Inhibition of HIV-1-induced Syncytia by 1,8-naphthalimide Photoactive Compound. T.C. Chanh, D.E. Lewis, J.S. Allan, F. Sogandares-Bernal, M.M. Judy, R.E. Utecht and J.L. Matthews. *AIDS Res & Human Retrovi* 9:891-896, 1993.

4-Alkylamino-3-bromo-N-alkyl-1,8-naphthalimides: New Photochemically Activatable Antiviral Compounds. S-C. Chang, B.J. Archer, R.E. Utecht, D.E. Lewis, M.M. Judy and J.L. Matthews. *Biomed Chem Letters* 3:555-556, 1993.

Photochemical Cross-linking of Proteins: Potential for Tissue Welding Without Heat. M.M. Judy, J.L. Matthews, R.L. Boriack, and A. Burlacu. *Soc for Optical and Quantum Electronics*, 1993.

Photoautomerization-S.E.T.: A New Oxygen-independent Mechanism for the Membrane-based Photochemical Inactivation of Enveloped Viruses. D.E. Lewis, R.E. Utecht, S-C. Chang, N.J. Umback, L.J. Costello, T.C. Chanh, M.M. Judy, J.T. Newman and J.L. Matthews. *The Spectrum*, 4:8-14, 1993.

XeCl Laser Ablation of Atherosclerotic Aorta: Luminescence Spectroscopy of Ablation

Products. A. Oraevsky, G.H. Petit, S.L. Jacques, R.A. Sauerbrey, F.K. Tittel, P.D. Henry. *Lasers Surg Med* 13:168-178, 1993.

The Role of Tissue Optics and Pulse Duration on Tissue Effects During High-power Laser

Irradiation. S.L. Jacques. *Applied Optics* , 32(13):2447-54, 1993.

Pulsed Photothermal Radiometry of Portwine Stain Lesions. S.L. Jacques, J.S. Nelson,

W.H. Wright and T.E. Milner. *Applied Optics* , 32(13): 2439-46, 1993.

Thermal Damage of Blood Vessels in a Rat Skin Flap Window Chamber Using

Indocyanine Green and a Pulsed Alexandrite Laser. S.T. Flock, S.L. Jacques.  
*Lasers in Med Sci* . 8(3):185-196, 1993.

Soft Tissue Studies with 805 nm Diode Laser Radiation; Thermal Effects with Contact Tips

and Comparison with Effects of 1064 nm Nd:YAG Laser Radiation. M.M. Judy,  
J.L. Matthews, B.L. Aronoff. *Lasers Surg Med* 13:528-532, 1993.

Effects of Preactivated MC540 in the Treatment of Lymphocytic Plasmacytic Stomatitis in

Feline Leukemia Virus and Feline Immunodeficiency Virus Positive Cats. *J Vet Dent*  
10:9-13, 1993.

Biodistribution and Toxicity of Photoproducts of Merocyanine 540. S. Pervaiz, M.

Battaglini, J.L. Matthews, *Cancer Chemo and Pharmacol* 31:467-474, 1993.

*In vitro* and *in vivo* Growth Suppression of Cancer-7 Human Breast Cancer by Novel

Photoproducts and Tamoxifen. K.S. Gulliya, R.K. Sharma, J.L. Matthews, A.C. Benniston, A. Harriman and J. Nemunaitis. *Cancer* 74:1725-1732, 1994.

Topoisomerase II Dependent Novel Antitumor Compounds Merocil and Merodantoin

Induce Apoptosis in Daudi Cells. K.S. Gulliya, B. Franck, U. Schneider, R. Sharma, L. Arnold and J.L. Matthews. *Anti-Cancer Drugs*. 5:557-566, 1994.

Preclinical Studies with Preactivated Compounds. K. Gulliya, R. Sharma, B. Wiggs, L.

Matthews, B. Franck in Biologic Effects of Light. E. Jung and M. Holick (eds) Walter de Gryter, Berlin 349-362, 1994.

Inhibition of Retrovirus-induced Syncytium Formation by Photoproducts of a Brominated

1-8,naphthalimide Compound. T. Chanh, D.E. Lewis, M.M. Judy, F. Sogandares-Bernal, G. Michalek, R. Utecht, H. Skiles, J.L. Matthews. *Antiviral Research* 25:133-146, 1994.

Biomedical Lasers in: Handbook of Biomedical Engineering, M.M. Judy, CRC Press Inc.

Capt. 87, pp. 1333-1345, 1995.

Cross-linking of Viral Envelop Proteins with 108, naphthalimide: A Major Mechanism for

Viral Inactivation. Y. Yang, H. Skiles, J.T. Newman, M.M. Judy, J.L. Matthews. *Proc of Jiangsu Chinese Med Research Inst*, 1995.

Photodynamic Inactivation of Viruses. J.L. Matthews and M.M. Judy in: Molecular

Biology and Biotechnology, R.A. Meyers (ed) VCH Publishers, New York, NY, pp 953-957, 1995.

Preliminary Results on Reflectance Feedback Control of Photocoagulation *in vivo*. M.R.

Jerath, R. Chandru, S.F. Barrett, H.G. Rylander III, A.J. Welch. *IEEE Trans Biomed Eng*, 41:201-203, 1994.

Digital Tracking and Control of Retinal Image. S.F. Barrett, M.R. Jerath, H.G. Rylander III, A.J. Welch. *Optical Eng* 41:201-203, 1994.

Temperature Control During Laser Vessel Welding. T.A. Springer, A.J. Welch. *Appl Optics*, 32(4):517-525, 1993.

Controlled Temperature Tissue Fusion: Argon Lesion Welding of Canine Intestine *in vitro*. I.F. Celisiz, T. Springer, S. Thomsen, A.J. Welch. *Lasers Surg Med* (in press).

Thermal Feedback Control During Laser-assisted Tissue Welding. I.F. Celisiz (PhD dissertation), The University of Texas at Austin, December, 1994.